

Manuel Kleiner  
www.kleinerlab.org

North Carolina State University  
Dept. of Plant & Microbial Biology  
Campus Box 7612  
4510B Thomas Hall  
Raleigh, NC 27695

+1 919 515 3792 (office)  
+1 812 391 2577 (mobile)  
manuel\_kleiner@ncsu.edu  
Twitter: @Manuel\_Kleiner

## RESEARCH AND TEACHING INTERESTS

Microbial physiology and metabolism, symbiosis, intra- and interspecies microbe-microbe interactions, genomics and metagenomics, proteomics and metaproteomics, environmental microbiology, evolution, horizontal gene transfer, the role of bacteriophages in horizontal gene transfer, microbial ecology, marine microbiology, renewable resources, and origins of life.

## CURRENT POSITION

8/2017      Assistant Professor, Chancellor's Faculty Excellence Program Cluster on  
Microbiomes and Complex Microbial Communities, NC State University

## EDUCATION AND PROFESSIONAL EXPERIENCE

- 9/2014 –      Banting **Postdoctoral** Fellow, University of Calgary  
8/2017      Energy Bioengineering Group, Marc Strous (advisor)
- 1/2013 –      **Guest Faculty**, Lora Hooper's Laboratory, UT Southwestern Medical Center, Dallas  
8/2017
- 1/2013 –      **Postdoctoral** Scientist, Max Planck Institute for Marine Microbiology, Bremen,  
8/2014      Germany
- 2012      **Ph.D.**, summa cum laude, Max Planck Institute for Marine Microbiology, Bremen,  
Germany  
Symbiosis Group, Nicole Dubilier (advisor)  
Dissertation title: "Metabolism and evolutionary ecology of chemosynthetic  
symbionts from marine invertebrates."
- 2008/2009      Postgraduate research year: Visiting fellow in the labs of James Reilly, Indiana  
University, Bloomington, IN and Nathan VerBerkmoes, Oak Ridge National  
Laboratory, Oak Ridge, TN
- 2008      **Diplom** in Biology, University of Greifswald, Germany (4 years of classes + 1 year  
thesis work in the groups of Michael Hecker and Thomas Schweder)  
Major: Microbial Physiology / Molecular Biology  
Minors: Microbial Ecology and Computer Science

Thesis title: “Metaproteomics of the bacterial symbionts of the marine oligochaete worm *Olavius algarvensis*.”

## PUBLICATIONS

(\* = contributed equally, ^ = corresponding authorship, \$ = student supervised by MK)

- Rubin-Blum, M., N. Dubilier and **M. Kleiner**<sup>^</sup> (2019). Genetic evidence for two carbon fixation pathways (the Calvin-Benson-Bassham Cycle and the Reverse Tricarboxylic Acid Cycle) in symbiotic and free-living bacteria. *mSphere* 4:e00394-18.
- Duerkop, B. A.\*, **M. Kleiner**<sup>\*^</sup>, D. Paez-Espino, W. Zhu, B. Hassell, S. E. Winter et al. (2018). Murine colitis reveals a disease-associated bacteriophage community. *Nature Microbiology* 3: 1023–1031.
- Kleiner, M**<sup>^</sup>, X. Dong, T. Hinzke\$, J. Wippler\$, E. Thorson\$, B. Mayer and M. Strous (2018). Metaproteomics method to determine carbon sources and assimilation pathways of species in microbial communities. *Proceedings of the National Academy of Sciences* 115(24): E5576-E5584.
- Zorz, J. K.\$, J. A. Kozlowski, L. Y. Stein, M. Strous and **M. Kleiner**<sup>^</sup>(2018). Comparative proteomics of three species of ammonia-oxidizing bacteria. *Frontiers in Microbiology* 9(938).
- Fida, T. T., J. Voordouw, M. Ataeian, **M. Kleiner**, G. Okpala, J. Mand and G. Voordouw (2018). Synergy of sodium nitroprusside and nitrate in inhibiting the activity of sulfate reducing bacteria in oil-containing bioreactors. *Frontiers in Microbiology* 9(981).
- Hinzke T., **M. Kleiner** and S. Markert (2018). Centrifugation-based enrichment of bacterial cell populations for metaproteomic studies on bacteria-invertebrate symbioses. *Methods in Molecular Biology, Microbial Proteomics*. D. Becher, Ed. Springer. 1841: 319-334
- Kleiner, M**<sup>\*^</sup>, E. Thorson\*\$, C. E. Sharp, X. Dong, C. Li and M. Strous (2017). Assessing species biomass contributions in microbial communities via metaproteomics. *Nature Communications* 1558.
- Ponnudurai, R., L. Sayavedra, **M. Kleiner**, S. E. Heiden, A. Thürmer et al. (2017). Genome sequence of the sulfur-oxidizing *Bathymodiolus thermophilus* gill endosymbiont. *Standards in Genomic Sciences* 12:50.
- Rubin-Blum, M., [...], **M. Kleiner** et al. (2017). Short-chain alkanes fuel mussels and sponge *Cycloclasticus* symbionts from deep-sea gas and oil seeps. *Nature Microbiology* 2:17093.
- Dong, X., **M. Kleiner**, C. E. Sharp, E. Thorson\$, C. Li, D. Liu and M. Strous (2017). Fast and simple analysis of MiSeq amplicon sequencing data with MetaAmp. *Frontiers in Microbiology* 8(1461).
- Kleiner, M**<sup>^</sup> (2017). Normalization of metatranscriptomic and metaproteomic data for differential gene expression analyses: The importance of accounting for organism abundance. *PeerJ Preprints* 5:e2846v1.
- Ponnudurai, R. P.\*, **M. Kleiner**\*, L. Sayavedra, J. M. Petersen, M. Moche, A. Otto et al. (2017). Metabolic and physiological interdependencies in the *Bathymodiolus azoricus* symbiosis. *The ISME Journal* 11: 463-477.

- Wippler, J.\* \$, **M. Kleiner**\*^, C. Lott, A. Gruhl, P. Abraham, R. Giannone et al. (2016). Transcriptomic and proteomic insights into innate immunity and adaptations to a symbiotic lifestyle in the gutless marine worm *Olavius algarvensis*. BMC Genomics 17(1): 942.
- Yu, Y.-T. N., **M. Kleiner** and G. J. Velicer (2016). Spontaneous reversions of an evolutionary trait loss reveal regulators of a sRNA that controls multicellular development in the myxobacteria. Journal of Bacteriology 198(23): 3142-3151.
- Petersen, J. M., A. Kemper, H. Gruber-Vodicka, U. Cardini, M. van der Geest, **M. Kleiner** et al. (2016). Chemosynthetic sulphur-oxidizing symbionts of marine invertebrate animals are capable of nitrogen fixation. Nature Microbiology 2:16195.
- Hamann, E., H. Gruber-Vodicka, **M. Kleiner**, H. Tegetmeyer, D. Riedel, S. Littmann et al. (2016). Environmental Breviatea harbor mutualistic *Arcobacter* epibionts. Nature 534: 254-258.
- Zimmermann, J., C. Wentrup, M. Sadowski, A. Blazejak, H. Gruber-Vodicka, **M. Kleiner** et al. (2016). Closely coupled evolutionary history of ecto- and endosymbionts from two distantly-related animal phyla. Molecular Ecology 25(13): 3203-3223.
- Schimak, M. P., **M. Kleiner**, S. Wetzel, M. Liebeke, N. Dubilier and B. Fuchs (2016). MiL-FISH: Multi-labelled oligonucleotides for fluorescence *in situ* hybridisation improve visualization of bacterial cells. Applied and Environmental Microbiology 82(1): 62-70.
- Kleiner, M.**\*^, C. Wentrup\*, T. Holler, G. Lavik, J. Harder, C. Lott, S. Littmann, M. M. M. Kuypers and N. Dubilier (2015). Use of carbon monoxide and hydrogen by a bacteria-animal symbiosis from seagrass sediments. Environmental Microbiology 17(12): 5023-5035.
- This paper was featured** on the BacterioFiles Podcast (#240)
- Sayavedra, L., **M. Kleiner**, R. Ponnudurai, S. Wetzel, E. Pelletier, V. Barbe et al. (2015). Abundant toxin-related genes in the genomes of beneficial symbionts from deep-sea hydrothermal vent mussels. eLife 4:e07966.
- Kleiner, M.**, L. V. Hooper and B. A. Duerkop (2015). Evaluation of methods to purify virus-like particles for metagenomic sequencing of intestinal viromes. BMC Genomics 16(7).
- Winkel M., P. Pjevac, **M. Kleiner**, S. Littmann, A. Meyerdierks, R. Amann and M. Mußmann (2014). Identification and activity of acetate-assimilating bacteria in diffuse fluids venting from deep-sea hydrothermal systems. FEMS Microbiology Ecology 90(3): 731-746.
- Kleiner, M.**\*^, J. C. Young\*, M. Shah, N. C. VerBerkmoes and N. Dubilier (2013). Metaproteomics reveals abundant transposase expression in mutualistic endosymbionts. mBio 4(3): e00223-13.
- Kleiner, M.**^, C. Wentrup, C. Lott, H. Teeling, S. Wetzel, J. Young et al. (2012). Metaproteomics of a gutless marine worm and its symbiotic microbial community reveal unusual pathways for carbon and energy use. Proceedings of the National Academy of Sciences 109(19): E1173-E1182.
- This paper was highlighted** in numerous newspapers, magazines and blogs e.g. Science Daily, CNN light years and in Nature.
- Kleiner, M.**^, J. M. Petersen and N. Dubilier (2012). Convergent and divergent evolution of metabolism in sulfur-oxidizing symbionts and the role of horizontal gene transfer. Current Opinion in Microbiology 15(5): 621-631.

**Kleiner, M.**, T. Woyke, C. Ruehland and N. Dubilier (2011). The *Olavius algarvensis* metagenome revisited: lessons learned from the analysis of the low diversity microbial consortium of a gutless marine worm. Handbook of Molecular Microbial Ecology II: Metagenomics in Different Habitats. F. J. d. Bruijn, Ed. Hoboken, NJ, USA, John Wiley & Sons, Inc. 2: 321-334.

Markert, S., A. Gardebrecht, H. Felbeck, S. M. Sievert, J. Klose, D. Becher, D. Albrecht, A. Thürmer, R. Daniel, **M. Kleiner**, M. Hecker and T. Schweder (2011). Status quo in physiological proteomics of the uncultured *Riftia pachyptila* endosymbiont. Proteomics 11(15): 3106-3117.

## MANUSCRIPTS IN REVIEW / REVISION

Hinzke, T., M. Strous and **M. Kleiner**<sup>^</sup> (submitted to Frontiers). More is not always better: Evaluation of 1D and 2D-LC-MS<sup>2</sup> methods for metaproteomics.

## MANUSCRIPTS IN PREPARATION

**Kleiner, M.**, C. Bergin, C. Lott, L. Polerecky, N. Musat, T. Vagner, M. M. M. Kuypers and N. Dubilier. “Single-cell analysis and modeling of autotrophic carbon fixation and carbon transfer in a gutless marine worm symbiosis.” (In preparation for submission to *The ISME Journal*)

Wippler, J., C. Wentrup, T. Woyke, N. Dubilier and **M. Kleiner**. “Ongoing sympatric speciation of an obligate bacteria-animal symbiosis in its Mediterranean habitat.” (In preparation for submission to *Nature Ecology & Evolution*)

**Kleiner, M.**, A. Bar-Even and M. Strous. “Comparative genomics of the bacterial Calvin cycle: variations in its genetic make up and its taxonomic distribution.” (In preparation for submission to *Nature Microbiology*)

H. R. Gruber-Vodicka, **Kleiner, M.**, C. Wentrup, N. Leisch, J. Ott and N. Dubilier. “*Candidatus* Thiosymbion spp., a genus of thiotrophic symbionts most closely related to the Chromatiaceae .” (In preparation for submission to *Systematic and Applied Microbiology*)

## AWARDS, HONORS AND RESEARCH SUPPORT

2019 – 2021	Foundation for Food and Agriculture Research (FFAR) New Innovator Award
2018	Plant Soil Microbial Community Consortium grant (PSMCC, \$60,000)
2015	Banting Postdoctoral Fellowship Award, NSERC (\$140,000; considered the most prestigious postdoctoral fellowship in Canada)
2014	Eyes High Postdoctoral Fellowship Award, University of Calgary (\$100,000)
2014	Friedrich Hirzebruch PhD thesis award of the German National Academic Foundation for outstanding work in the fields of Mathematics, Natural Sciences and Engineering (5000 Euro)
2013	Finalist for the Society in Science Branco Weiss Fellowship
2012	Sequencing grant from the Community Sequencing Program of the US Department of Energy Joint Genome Institute for the sequencing of 20 bacterial genomes and metagenomes (PI): “ <i>Understanding novel pathways for energy and carbon use in bacterial symbionts of gutless marine worms.</i> ”

- 2011 MarMic (International Max Planck Research School for Marine Microbiology) teaching excellence award (best tutorial)
- 2009 - 2012 PhD scholarship award from the German National Academic Foundation (Studienstiftung des deutschen Volkes) for academically gifted students
- 2009 Poster Award at the Gordon Research Conference for Applied and Environmental Microbiology
- 2008/09 Hölderlin-scholarship from the German National Academic Foundation for a postgraduate research year in the USA
- 2007 Travel scholarship from the German National Academic Foundation for a research internship in the Laboratory of Gregory Velicer, Indiana University
- 2005 Travel grant of the International Society for the Study of the Origin of Life (ISSOL) to attend its conference in Beijing
- 2005 - 2012 Scholarship from e-fellows.net supplying fast internet access for academically gifted students
- 2001 – 2008 Graduate scholarship award from the German National Academic Foundation (Studienstiftung des deutschen Volkes) for academically gifted students
- 1997/98 Scholarship from AFS Intercultural Programs to spend a year of high school in Portugal

## **SERVICE - DISCIPLINE**

4/2015 - **Associate Editor**, *Frontiers in Microbiology: Microbial Physiology and Metabolism*

**Reviewer activity:** *Molecular Systems Biology*, *mBio* (Guest Editor), *Environmental Microbiology*, *Marine Ecology*, *Nature Communications*, *Trends in Microbiology*, *PLoS Genetics*, *Science*, *ISME J*, *Genome Biology*, *Biological Reviews*, *PeerJ*, *Computational Structural Biotechnology Journal*, *Symbiosis* and *Czech Science Foundation (GACR)*, *Austrian Science Fund (FWF)*.

I am currently a member of six PhD Thesis Committees at NC State University, the University of Calgary, the Max Planck Institute in Bremen and the University of Greifswald.

## **CONFERENCES AND INVITED TALKS**

- Dec 2018 Invited Talk, International Metaproteomics Symposium, Leipzig, Germany
- Nov 2018 Invited Talk, UT Knoxville Microbiology Seminar
- Oct 2018 Invited Talk, Triangle Microbial Interactions Meeting, Cary, NC
- Aug 2018 Invited Talk, BASF, Durham, NC
- May 2018 Invited Talk, Center for Integrated Fungal Research, NC State
- May 2018 Talk on Viral Metagenomics, Microbiome Monthly Meetup, NC State
- Apr 2018 Invited talk, Novozymes, Durham, NC
- Apr 2018 Invited talk, Microbiome Seminar Series, UNC Chapel Hill, NC
- Mar 2018 Talk, DOE JGI Genomics of Energy and Environment Meeting, San Francisco
- Feb 2018 Invited talk, Ag Biotech Summit, NC Biotechnology Center, Chapel Hill, NC
- Dec 2017 Invited talk, Computational Biology Seminar Series, Veterinary Medicine, NC State
- Oct 2017 Invited talk, Biology Seminar Series, Georgia Institute of Technology
- Oct 2017 Invited talk, Marine Sciences Seminar Series, UNC Chapel Hill, NC
- Sep 2017 Invited talk, Biochemistry Seminar Series, NC State University
- Sep 2017 Invited talk, Dept. of Symbiosis, Max Planck Institute for Marine Microbiology

- 2017 Invited talk, ASM Microbe conference, New Orleans. “Assessing the metabolism, physiology and interspecies interactions in microbial symbioses via metaproteomics.”
- 2016 Talk, International Society for Microbial Ecology (ISME) conference, Montreal, Canada. “Assessing species biomass contributions in microbial communities via metaproteomics.”
- 2016 Attendee, DOE Joint Genome Institute User Meeting, Walnut Creek, CA
- May 2015 Invited talk, Seminar Series, Bermuda Institute of Ocean Sciences, Bermuda
- 2015 Poster, DOE Joint Genome Institute User Meeting, Walnut Creek, CA
- 2014 Poster, DOE Joint Genome Institute User Meeting, Walnut Creek, CA
- Feb 2014 Invited talk, Biology Seminar Series, Baylor University, TX
- 2013 Talk, American Society for Microbiology (ASM) General Meeting in Denver, Colorado. “Novel metabolic pathway for the massive assimilation of host waste products into carbon storage by the chemolithoautotrophic symbiont of a gutless marine worm.”
- 2012 Poster, Gordon Research Seminar and Conference on the Molecular Basis of Microbial One-Carbon Metabolism at the Bates College, ME, USA. “Proposal of a pyrophosphate-dependent, more energy efficient version of the Calvin cycle.”
- 2012 Poster, EU-US Environmental Biotechnology Workshop on Microbial Community Dynamics: Cooperation and Competition in St. Louis, MO, USA.
- 2011 Invited talk, MIMAS (Microbial interactions in marine systems) Symposium in Greifswald, Germany. “Metaproteomics of a gutless marine worm and its symbiotic microbial community: Discovery of novel pathways for carbon and energy use.”
- 2011 Attendee, NanoSIMS user meeting, Warnemünde, Germany
- 2010 Talk and poster, Symbiotic interactions meeting in Würzburg, Germany
- 2010 Talk, International Society for Microbial Ecology conference in Seattle, Washington, USA. “A symbiosis fueled by carbon monoxide: novel insights in the microbial community of a gutless marine worm.”
- 2009 Poster, Gordon Research Conference on Applied & Environmental Microbiology at the Mount Holyoke College, USA. “Metaproteomics of a gutless marine worm and its symbiotic microbial community.”
- 2009 Talk, International Symbiosis Society Congress in Madison, Wisconsin, USA. “Metaproteomics of a gutless marine worm and its symbiotic microbial community: Novel insights into host and symbiont metabolism and interactions.”

## **RESEARCH EXPERIENCE AND TRAINING**

- Fall 2018 ALLI – Agricultural Leadership Learning Institute, (8 sessions), NCSU
- Feb 2015 One day communication skills workshop, Mitacs Inc., University of Calgary
- Jan 2014 Two day science communication workshop, Klaus Tschira Foundation, Heidelberg
- Sep 2013 Microbial Genomics & Metagenomics workshop (one week), DOE Joint Genome Institute, Walnut Creek, California
- July 2013 Three day course on the R software environment for statistical computing and graphics
- Sep 2011 Two day grant application training workshop, Max Planck Institute
- Mar 2011 Two day scientific writing course, Max Planck Institute
- Feb 2011 Two day course on leadership skills for scientists, Max Planck Institute
- Dec 2010 Two weeks of training to become a NanoSIMS operator, Max Planck Institute
- Nov 2010 RAST workshop at the Argonne National Laboratory. ‘Genome annotation, comparison and metabolic modeling’

- May 2010 Biosand workshop 2010 at the Hydra Institute for Marine Sciences, Elba, Italy
- 2008/09 Postgraduate research year, Indiana University and Oak Ridge National Laboratory: Additional training in Proteomics and Mass spectrometry.
- 2007 Three months research internship in the group of Gregory Velicer, Indiana University. ‘Developing a mutant screen for *Myxococcus xanthus* showing special fruiting body development’.
- 2006/07 Three month research internship in the bioinformatics group at the Max Planck Institute for Marine Microbiology. ‘Developing software for combined analyses of proteome and genome data’.
- 2006 Two month research internship in the group of Michael Thomm at the University of Regensburg. ‘Heterologous expression of archaeal proteins in *Escherichia coli*’.
- 2006 Five week research internship at the Institute for Marine Biotechnology in Greifswald. ‘Proteomics of the *Riftia pachyptila* symbionts’.
- 2005 – 2007 Extracurricular ‘Life Sciences Academy’ of the Studienstiftung des dt. Volkes on Evolution and Development taught by Andrei Lupas, MPI Tübingen
- 2005/06 Research Assistant in the Molecular Biology Department of the University of Greifswald. ‘Analyzing codon usage in stress specific *Bacillus subtilis* proteins and developing methods for absolute protein quantification’.
- Sept 2004 Summer academy of the Studienstiftung des dt. Volkes “Extra solar planets and extraterrestrial life”.

## TEACHING EXPERIENCE

- Fall 2018 Taught three credit course on “Microbial Symbiosis & Microbiomes” (BSC495/MB590), NC State University
- Spring 2018 Taught guest lectures in Aquatic Microbiology (MEA 493) and Meet Your Microbes (LSC 170), NC State University
- Fall 2016 Taught workshop on Negotiation Skills in Geoscience Graduate Skills course (GLGY 699.56), University of Calgary
- Fall 2016 Taught lectures on the origins of life in a course for geologists on the “Evolution of Earth through Life and Time” (GLGY 493), University of Calgary
- Spring 2016 Taught lectures on the origins of life in a course for geologists on the “Evolution of Earth through Life and Time” (GLGY 493), University of Calgary
- Fall 2015 Taught part of Microbial Physiology course (CMMB 443), University of Calgary
- 2014 Teaching faculty at the SYMBIOMICS field workshop on Elba, Italy. Mix of lectures, seminars and laboratories for a group of 20 international MSc and PhD students (two weeks).
- 2012 Teaching faculty at the SYMBIOMICS field workshop on Elba, Italy (two weeks).
- 2011 Instructor for tutorials accompanying lectures on metagenomics (Group of 20 MSc students)
- 2011 Design of and sole instructor for a two day workshop on ‘Genome annotation, comparison and metabolic modeling using RAST, the SEED and Model-SEED’ for a group of 15 graduate students and postdocs. The workshop consisted of lectures, demonstrations and hands on exercises. I received teaching evaluations above faculty average.
- 2011 Instructor for tutorials accompanying lectures on symbiosis for 20 MSc students. I received the MarMic teaching excellence award for the best tutorial.

## INDIVIDUAL GUIDANCE AND MENTORSHIP OF STUDENTS AND POSTDOCS

- 2019 – pres. Undergraduate research student (Mariska Thayagan), Kleiner Lab, NCSU
- 2018 – pres. MS student (Natalia Marino), Kleiner Lab, NCSU
- 2018 – pres. MS student (Alexandria Bartlett), Kleiner Lab, NCSU
- 2018 – pres. Undergraduate research student (Rebekah Lim), Kleiner Lab, NCSU
- 2018 – pres. Undergraduate research student (Amani Albrecht), Kleiner Lab, NCSU
- 2018 – pres. PhD student (Angie Mordant), Kleiner Lab, NCSU
- 2018 – pres. Postdoc (Fernanda Salvato), Kleiner Lab, NCSU
- 2018 – pres. PhD student (Nick Taylor), Kleiner Lab, NCSU
- 2017 – pres. Undergraduate research student (Jessie Maier), Kleiner Lab, NCSU
- 2017 Six months, visiting PhD student (Tjorven Hinzke), Energy Bioengineering Group, University of Calgary
- 2016 – pres. PhD student (Jackie Zorz), Energy Bioengineering Group, University of Calgary
- 2016/17 Project student (Erin Thorson), Energy Bioengineering Group, University of Calgary
- 2016 Four months summer student (Erin Thorson), Energy Bioengineering Group, University of Calgary
- 2011 - 2016 PhD student (Juliane Wippler), Department of Symbiosis, Max Planck Institute for Marine Microbiology (PhD received in 2016)
- 2011 Guest PhD student (Ruby Ponnudurai) doing a 4 week project, Symbiosis Group, Max Planck Institute for Marine Microbiology
- 2011 Six week Lab Rotation MSc student (Lizbeth Sayavedra), Symbiosis Group, Max Planck Institute for Marine Microbiology
- 2010/11 MSc student (Juliane Wippler), Symbiosis Group, Max Planck Institute for Marine Microbiology
- 2010 Research intern for 8 weeks (MSc student Johannes Boetschi from the University of Greifswald), Symbiosis Group, Max Planck Institute for Marine Microbiology

## FIELD EXPERIENCE

- 2018 NCSU Center for Marine Sciences and Technology, North Carolina. Collection of chemosynthetic symbioses from marine sediments (multiple trips).
- 2015 Bermuda Institute of Ocean Sciences, Bermuda. Collection of marine sediments and symbionts from gutless marine oligochaetes for metagenomics, metatranscriptomics and metaproteomics.
- 2013 Little Darby Island, Bahamas. Collection of marine sediment and symbiotic Meiofauna for metagenomics and metaproteomics.
- 2012\* Heron Island Research Station, Australia. Collection of symbiotic Meiofauna for metagenomics and metaproteomics.
- 2012 Hydra Institute for Marine Sciences, Elba, Italy. Collection of gutless oligochaetes, *in situ* measurements and physiological incubation experiments.
- 2011\* Hydra Institute for Marine Sciences, Elba, Italy. Collection of gutless oligochaetes.
- 2010\* Hydra Institute for Marine Sciences, Elba, Italy (Two Trips). Collection of gutless oligochaetes, *in situ* measurements and physiological incubation experiments.
- 2009\* Bermuda Institute of Ocean Sciences, Bermuda. Collection of marine sediment and symbionts from gutless marine oligochaetes for metaproteomics and metagenomics.
- 2007\* Hydra Institute for Marine Sciences, Elba, Italy. Collection of symbionts from gutless marine oligochaetes for metaproteomics.



\*For these sampling trips I was the main coordinator of the research team - consisting of three to five scientists - and was in charge of equipment and sample logistics.

## **SKILLS AND EXPERTISE**

Microbial Ecology, Microbiology, Microbial Metabolism and Physiology, Symbiosis, Marine Microbiology, Biochemistry, Bioinformatics, Environmental Microbiology, Bacterial Genetics, Zoology, Evolution and Development, Genomics and Proteomics, Metagenomics and Metaproteomics and Mass Spectrometry (including NanoSIMS).

### **Basic Microbiology and Molecular Biology Skills:**

DNA extraction, PCR, qPCR, cloning, Sanger sequencing, fluorescence *in situ* hybridization (FISH, CARD-FISH), (fluorescence) microscopy, cultivation of bacteria, cultivation of bacteriophages, anaerobic work, and Western blots.

### **Specialized Skills:**

Bioinformatics (command line scripting, basic programming, database design and setup, handling of large scale datasets and extensive experience with a large number of bioinformatics tools), experienced NanoSIMS operator, genomics and metagenomics (PacBio and Illumina data: read QC, assembly, binning, annotation, metabolic and phylogenetic reconstruction and comparative genomics), proteomics and metaproteomics (protein and peptide separations, operation and maintenance of various mass spectrometers, protein identification, peptide de novo sequencing, data analysis, statistics and interpretation), enrichment of specific bacteria from mixtures using a variety of centrifugations and filtrations, bacteriophage and virus purification using ultracentrifugation, fluorescence-activated cell sorting (FACS), multiple displacement amplification (MDA), operation of a variety of HPLC and GC instruments, GC-MS and LC-MS, confocal laser scanning microscopy, and metabolomics data analyses.

### **Transferable skills relevant to science:**

Coordination of scientific projects with multiple international collaboration partners, training in conflict management (trained through volunteer work for large intercultural exchange organization), diving license, experience in underwater sampling in marine sediments, licenses for sailing and motor boats, construction of photovoltaic power generators.

## **PROFESSIONAL MEMBERSHIPS**

- 2009 - International Symbiosis Society
- 2010 - International Society for Microbial Ecology
- 2013 - American Society for Microbiology (ASM)